# Evaluation of CYP3A activity in humans using three different parameters based on endogenous cortisol metabolism 

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#### Abstract

Aim: Currently, there is considerable debate as to which method is more accurate for measuring the activity of CYP3A in vivo: cortisol $6 \beta$-hydroxylation clearance ( $\left(l_{m(6 \beta)}\right)$ or the urinary ratio of $6 \beta-O H F$ to $F(6 \beta-O H F / F)$. Furthermore, the value of measuring endogenous levels of cortisol over a 24 h period $\left(\mathrm{AUC}_{F}\right)$ needs to be confirmed. The aim of the present study was to determine which method was most effective at measuring changes in the in vivo activity of CYP3A: $A \cup C_{F}, C l_{m(6 \beta)}$, or $6 \beta-0 H F / F$. Methods: A two phase, cross-over design was adopted in this study. A total of 24 subjects ( 12 males and 12 females) were randomly assigned to one of two groups: the test group subjects were given 250 mg clarithromycin tablets twice a day for a period of 4 d , whereas the control group received a placebo twice daily for a similar period. On d 5 of the study, the last dose of either clarithromycin or placebo was supplemented with an oral dose of 7.5 mg midazolam (MDZ); blood and urine samples were then collected at various times. All samples collected at the same sampling times on d 4 were used to evaluate the effects of MDZ administration on cortisol levels and metabolism. The ratio of 1-hydroxymidazolam (1-OHMDZ) concentration to MDZ concentration at 1 h (MR) was taken as a measure of the in vivo CYP3A activity. $\mathrm{AUC}_{\mathrm{F}}, \mathrm{Cl}_{\mathrm{m}(6 \beta)}$, and $6 \beta-\mathrm{OHF} / \mathrm{F}$ were also used as biomarkers for CYP3A activity. Results: No correlations were found (either before or after inhibition) between CYP3A activity and any of the following measures: AUC $_{F}$, $\mathrm{Cl}_{\mathrm{m}(6 \beta)}$, or $6 \beta-\mathrm{OHF} / \mathrm{F}(r<0.4, P>0.05)$. After 4 d of clarithromycin administration, CYP3A activity (MR) decreased by $75 \%(P=0.000)$, whereas $A \cup C_{F}$ increased by $19 \%(P=0.040)$, and $C_{m(6 \beta)}$ and $6 \beta-0 H F / F$ decreased by $54.2 \%(P=0.000)$ and $50 \%(P=0.003)$, respectively. No significant changes in $\operatorname{AUC}_{F}(P=0.178)$, or in the amount of urinary $6 \beta-O H F(P=0.169)$, or in $F(P=0.391)$ were found over a 24 h time period, either with or without MDZ administration. Conclusion: Although $C_{m(6 \beta)}$ and $6 \beta-0 H F / F$ can reflect the decline in CYP3A activity, the impression they provide is neither accurate nor complete. $A^{\prime} C_{F}$ is completely ineffective for evaluating variations in CYP3A activity. MDZ administration had no evident effects on either cortisol metabolism or excretion over a period of 24 h .


Keywords: CYP3A activity; cortisol; 6ß-hydroxycortisol; midazolam
Acta Pharmacologica Sinica (2009) 30: 1323-1329; doi: 10.1038/aps.2009.116; published online 24 August 2009

## Introduction

Cytochrome P450 3A (CYP3A), which is highly expressed in the liver and in the intestine, is the most abundant cytochrome isoform in humans. CYP3A accounts for approximately $60 \%$ of the total cytochrome content in the liver. Nearly half of all current clinical drugs are substrates for CYP3A ${ }^{[1,2]}$. Overall, the activity of the CYP3A subfamily in adults is comprised mostly of CYP3A4 and CYP3A5 ${ }^{[2]}$. Because of the wide variability in CYP3A activity, it is necessary to study the CYP3A phenotype in order to predict the optimal dosage range of

[^0]drugs (ie, that which can improve therapeutic effects but minimize adverse effects $)^{[3,4]}$.
Several assessments of the CYP3A phenotype have been carried out in vivo; these include midazolam (MDZ) plasma clearance, the erythromycin breath test and the $6 \beta$-hydroxycortisol/ free cortisol $(6 \beta-\mathrm{OHF} / \mathrm{F})$ ratio in urine. Of these markers, the most widely used probe drugs are MDZ and erythromycin; however these biomarkers all have limitations.
MDZ is biotransformed primarily to 1-hydroxy-midazolam (1-OHMDZ) in vivo. This procedure is catalyzed by CYP3A enzymes. Both intestinal and hepatic CYP3A activity can be characterized by MDZ oral clearance, and this is typically used as a reliable index for CYP3A phenotyping ${ }^{[5]}$. However, in order to measure the oral clearance of MDZ, several blood
samples are often required. Zhu et al ${ }^{[6]}$ used single plasma sampling to predict CYP3A activity in ten healthy male volunteers in vivo. A significant correlation was demonstrated between weight-normalized MDZ oral clearance and the plasma concentration ratio of 1-OHMDZ to MDZ (MR) that was measured 1 h after intake of a single ( 7.5 mg ) oral dose of MDZ ( $r=0.7, P<0.05$ ). However, due to its pharmacodynamic activity as a central nervous system depressant, this technique is not convenient for the evaluation of CYP3A activity.
The intravenous administration of stable isotopically labeled erythromycin in the erythromycin breath test also has its limitations. In particular, this technique is capable of measuring only hepatic CYP3A activity.
The measurement of endogenous cortisol (F) levels may represent a safe, simple and non-invasive assay of CYP3A activity. It has been suggested that the urinary ratio of $6 \beta-\mathrm{OHF} / \mathrm{F}$ is also a useful marker of both the induction and the inhibition of hepatic CYP3A activity ${ }^{[7]}$. However, this measure does not appear to be an accurate marker of the pharmacokinetic properties of other substrates of CYP3A, such as the well-known probe drug MDZ and erythromycin ${ }^{[8-10]}$.
Recently, Furuta et al ${ }^{[11]}$ found that cortisol $6 \beta$-hydroxylation clearance $\left(\mathrm{Cl}_{\mathrm{m}(6 \beta)}\right)$ could be a reliable index for in vivo CYP3A activity evaluation. In this study, a strong correlation between endogenous and exogenous $6 \beta$-hydroxylation clearance was shown in three healthy male volunteers. Furthermore, when the macrolide antibiotic clarithromycin was administered to another healthy male volunteer, the inhibitory effects of clarithromycin on in vivo CYP3A activity were clearly indicated by the $6 \beta$-hydroxylation clearance of endogenous cortisol, but not by the urinary $6 \beta-\mathrm{OHF} / \mathrm{F}$. It has also been suggested that urinary $6 \beta-\mathrm{OHF} / \mathrm{F}$ does not always reflect the in vivo CYP3A activity ${ }^{[11]}$. However, significant shortcomings were associated with this study by Furuta et al; these included the limited number of samples, and the lack of a comparison with probes such as MDZ and erythromycin.

In these studies, the relationship between the endogenous cortisol $6 \beta$-hydroxylation clearance and the in vivo CYP3A activity (as reflected by the oral clearance of MDZ) was examined. We have previously published the results obtained using a high-performance liquid chromatography with ultraviolet absorbance detection (HPLC-UV) method for the simultaneous determination of $6 \beta$-OHF and F in human urine or plasma ${ }^{[12]}$. Of the 12 subjects ( 6 males and 6 females) enrolled in our study, a good correlation between cortisol $6 \beta$-hydroxylation clearance and MDZ oral clearance was seen in only 6 male subjects. The unexpected outcome of this study was the discovery of a negative correlation between the ratio of the area under the plasma concentration-time curve of 1-OHMDZ to that of MDZ $\left(\mathrm{AUC}_{\mathrm{MR}}\right)$ and the area under the plasma concentration-time curve of cortisol $\left(\mathrm{AUC}_{(0-t) \mathrm{F}}\right)$. A strong correlation between plasma cortisol concentrations at 1, $4,8,10$, and 24 h and $\mathrm{AUC}_{(0-24) \mathrm{F}}$ was also determined by linear regression. Based on these earlier findings, we proposed that the measure of $\mathrm{AUC}_{\mathrm{F}}$ that was calculated from plasma cortisol concentrations at $1,4,8,10$, and 24 h , represented a new bio-
marker for assessing in vivo CYP3A activity.
In conclusion, as the utility of $C l_{\mathrm{m}(6 \beta)}$ and $6 \beta-\mathrm{OHF} / \mathrm{F}$ measures were still up for debate, our laboratory sought to determine the effectiveness of $\mathrm{AUC}_{\mathrm{F}}$. Thus, the present study was undertaken to demonstrate whether $\mathrm{AUC}_{\mathrm{F}}, C l_{\mathrm{m}(6 \beta)}$, and $6 \beta-\mathrm{OHF} / \mathrm{F}$ provided accurate measures of variations in CYP3A activity in vivo. In addition, the effect of MDZ administration on cortisol metabolism and excretion over a 24 h time period was also investigated.

## Materials and methods

## Chemicals and reagents

MDZ, 1-hydroxy-midazolam, cortisol and 6 6 -hydroxycortisol (at least $98 \%$ purity) were purchased from Sigma-Aldrich. Acetonitrile and methanol were of HPLC grade. All other chemicals were of AR grade and were available from commercial sources.

## Subjects

Twenty-four normal volunteers ( 12 males, 12 females; age: $22.1 \pm 1.2$ years; weight: $58.6 \pm 5.4 \mathrm{~kg}$; height: $165.9 \pm 7.0 \mathrm{~cm}$ ) participated in this study. As indicated by their medical history, physical examinations, routine laboratory tests (hematology, blood chemistries and urinalysis), all subjects were in good health. The subjects were asked to abstain from the consumption of alcohol, caffeine and grapefruit juice for two weeks prior to the study. All of the subjects were non-smokers and consumed a normal diet. This experimental protocol was approved by the Ethical Committee of the School of Pharmaceutical Sciences, Central South University. Prior to commencing the study, written informed consent was obtained from each subject.

## Experimental protocol

A two-way crossover design was adopted in this test. A total of 24 volunteers ( 12 males and 12 females) were randomly assigned to one of two groups. After an overnight fast, either 250 mg clarithromycin tablets (experimental group) (Huiren, China) or placebo (control group) were given twice daily (at 08:00 and 20:00) to each subject, for a period of 4 d . On d 5 , the last dose of either clarithromycin or placebo was given with an oral dose of 7.5 mg MDZ (Roche, China). Blood samples were then collected both prior to and at $1,4,8,10,24 \mathrm{~h}$ after drug administration. Urine samples were collected at $0-4,4-8$, $8-10,10-24 \mathrm{~h}$ time periods. To evaluate the effects of MDZ administration on cortisol levels and metabolism, the samples that were collected on d 4 were compared with those obtained on the day of MDZ treatment. With the exception of water intake, food was prohibited for 2 h after MDZ administration. To separate the plasma, blood samples were centrifuged at $3500 \times g$ for 10 min as the volume of urine was recorded. The plasma and urine samples were stored at $-20^{\circ} \mathrm{C}$ until required for analysis. After a washout period of two weeks, two groups switched treatments respectively. Blood and urine samples were collected in the same way.

## Analytical methods

## $6 \beta-0 H F$ and $F$ in plasma or urine

$F$ and $6 \beta-\mathrm{OHF}$ in urine or plasma were measured using the HPLC-UV method that was developed in our laboratory ${ }^{[12]}$. The lower limit of quantitation was $6.9 \mu \mathrm{~g} / \mathrm{L}$ for $6 \beta-\mathrm{OHF}$ and $2 \mu \mathrm{~g} / \mathrm{L}$ for F in urine. The lower limit of quantitation was 7.8 $\mu \mathrm{g} / \mathrm{L}$ for F in plasma. The accuracy, determined for three different concentrations, ranged between $95 \%$ and $107 \%$.

## MDZ and 1-OHMDZ in plasma

MDZ and 1-OHMDZ in plasma were determined using a previously established HPLC method with minor modifications ${ }^{[13]}$. Briefly, to act as the internal standard, a solution of chlordiazeposide was added to 1.0 mL of human plasma. After the addition of $100 \mu \mathrm{~L}$ of $1.0 \mathrm{~mol} / \mathrm{L}$ sodium hydroxide, the plasma samples were extracted with 5.0 mL of cyclohexane/diethyl ether (3:7). HPLC analysis was performed using a Shimadzu LC-2010CHT high-performance liquid chromatography apparatus (Shimadzu, Kyoto, Japan). The separation of MDZ and 1-OHMDZ was achieved on a Hypersil GOLD column (4.6 $\mathrm{mm} \times 150 \mathrm{~mm}, 5 \mu \mathrm{~m}$, Thermo Electron, USA) using a mobile phase of methanol $/ 10 \mathrm{mmol} \cdot \mathrm{L}^{-1}$ phosphate buffer ( pH 7.4 ) $(57: 43, v / v)$. The flow rate was $1.0 \mathrm{~mL} / \mathrm{min}$ and the ultraviolet absorbance was monitored at 254 nm . The lower limit of quantitation was $2 \mu \mathrm{~g} / \mathrm{L}$ for 1-OHMDZ and $4 \mu \mathrm{~g} / \mathrm{L}$ for MDZ. The coefficients of variation for both intra-day and inter-day precision for each compound were less than $3.9 \%$.

## Data analysis

The plasma concentration ratio of 1-OHMDZ to MDZ (MR) at 1 h was used as a measure of the in vivo CYP3A activity. $\mathrm{AUC}_{\mathrm{F}}$ at each sample time was calculated from the plasma cortisol concentrations using a trapezoid ruler. $C l_{\mathrm{m}(6 \beta)}$ was calculated from the amount of urinary excreted $6 \beta$-OHF over a period of 24 h , divided by $\mathrm{AUC}_{\mathrm{F}}$. $6 \beta-\mathrm{OHF} / \mathrm{F}$ was calculated from the ratio of urinary $6 \beta-\mathrm{OHF}$ to F .

Parameter values obtained in the same subjects under different conditions were compared using Wilcoxon's rank sum test, whereas $\mathrm{AUC}_{\mathrm{F}}$, both before and after inhibition, was logarithmically transformed, and then evaluated using the variance analysis method. Correlations between $M R$ and $A U C_{F}$, between MR and $C l_{\mathrm{m}(6 \beta) \text {, }}$, and between MR and $6 \beta-\mathrm{OHF} / \mathrm{F}$ were determined using Spearman's rank correlation analysis. All statistical analyses were performed using SPSS 12.0 statistics software, and $P$ values less than 0.05 were considered to be significant.

## Results

## CYP3A activity variation with clarithromycin pretreatment

The in vivo CYP3A activity, reflected by MR, was reduced by $75 \% ~(~ P=0.000)$ following clarithromycin pretreatment. The significant change in MR is shown in Figure 1. Plasma MDZ, 1-OHMDZ concentration and CYP3A activity (MR) both before and after inhibition by clarithromycin are included in Table 1.


Figure 1. Variations in CYP3A activity (MR) before and after inhibition by clarithromycin ( $n=24$ ). MR: the plasma concentration ratio of 1-OHMDZ to MDZ at 1 h .

## AUC $_{F}$ variation and correlation with CYP3A activity

MDZ administration resulted in a $25 \%$ decrease ( $P=0.033$ ) in plasma cortisol concentrations at 1 h after MDZ administration, but there were no changes in concentration at other sampling times. $\mathrm{AUC}_{\mathrm{F}}$ was increased by $19 \%(P=0.040)$ when CYP3A activity was inhibited by clarithromycin (Table 2 and Figure 2). No correlations were found between $A U C_{F}$ and CYP3A activity either before or after inhibition $(r=0.342$, $P=0.102$; $r=0.391, P=0.059$; Figure 3). There was also no correlation between the change in $\mathrm{AUC}_{\mathrm{F}}$ and CYP3A activity ( $r=0.260, P=0.220$ ) (Figure 3).

## $\mathrm{Cl}_{\mathrm{m}(6 \beta)}$ variation and correlation with CYP3A activity

MDZ administration caused increases in $6 \beta-$ OHF urinary excretion of $51 \%(P=0.022), 75 \% ~(P=0.002)$ and $103 \% ~(P=0.019)$ at $0-4,4-8$, and $8-10 \mathrm{~h}$ time-periods, respectively. However, there were no increases in excretion and no alterations in $C l_{\mathrm{m}(6 \beta)}$ at 10-24 and 0-24 h time-periods. Following clarithromycin administration, there was a decrease in $6 \beta$-OHF urinary excretion at all time-periods, and a reduction in $C l_{\mathrm{m}(6 \beta)}$ of $54.2 \%$ ( $P=0.022$; Table 3 and Figure 2). However, there were no correlations found between $C l_{\mathrm{m}(6 \beta)}$ and MR, either before or after

Table 1. Mean values of plasma 1-hydroxy-midazolam, midazolam concentrations, and CYP3A activity (MR) before and after inhibition by clarithromycin. $n=24$. Data are given as mean $\pm$ SD.

| Time (h) | 1-OHMDZ ( $\mu \mathrm{g} / \mathrm{L}$ ) |  | MDZ ( $\mu \mathrm{g} / \mathrm{L}$ ) |  | MR |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Before inhibition | After inhibition | Before inhibition | After inhibition | Before inhibition | After inhibition |
| 1 | $12.38 \pm 4.47$ | $10.36 \pm 4.49$ | $26.96 \pm 8.82$ | $95.25 \pm 52.73$ | $0.49 \pm 0.19$ | $0.13 \pm 0.06$ |

MR: the plasma concentration ratio of 1-OHMDZ to MDZ at 1 h .

Table 2. Plasma cortisol concentration variations before and after clarithromycin administration. $n=24$. Data are given as mean $\pm$ SD.

| Time (h) | Cortisol concentration ( $\mu \mathrm{g} / \mathrm{L})$ <br> Before inhibition |  | After inhibition |
| :---: | :---: | :---: | ---: |
|  | Non MDZ | With MDZ |  |
|  |  |  |  |
| 1 | $77.26 \pm 40.10$ | $57.90 \pm 18.63$ | $61.35 \pm 22.96$ |
| 4 | $44.46 \pm 33.84$ | $50.28 \pm 17.40$ | $55.98 \pm 27.92$ |
| 8 | $46.48 \pm 37.56$ | $50.35 \pm 18.55$ | $61.65 \pm 28.02$ |
| 10 | $41.36 \pm 18.13$ | $46.13 \pm 14.57$ | $57.29 \pm 23.34$ |
| 24 | $97.02 \pm 49.23$ | $113.90 \pm 32.45$ | $136.72 \pm 48.62$ |
| AUC $_{\mathrm{F}}\left(\mu \mathrm{g} \cdot \mathrm{h} \cdot \mathrm{L}^{-1}\right)$ | $1459.56 \pm 696.24$ | $1609.17 \pm 357.89$ | $1918.97 \pm 620.25$ |



Figure 2. $A U C_{F}, C l_{m(6 \beta)}$, and $6 \beta-O H F / F$ variations during the $0-24 \mathrm{~h}$ timeperiod before and after clarithromycin pretreatment, and also in the absence of MDZ administration $(n=24) .{ }^{b} P<0.05,{ }^{c} P<0.01$. MR: the plasma concentration ratio of 1-OHMDZ to MDZ at 1 h .
inhibition ( $r=0.107, P=0.619 ; r=-0.317, P=0.132$, respectively; Figure 4). There was also a lack of a correlation between the change in $C l_{\mathrm{m}(6 \beta)}$ and MR ( $r=0.019, P=0.931$; Figure 4).

Table 3. Variations in $6 \beta-0 H F$ urinary excretion before and after clarithromycin inhibition. $n=24$. Data are given as mean $\pm$ SD.

| Time <br> period (h) | $6 \beta-$ OHF urinary excretion ( $\mu \mathrm{g}$ ) <br> Before inhibition <br> With MDZ |  | After inhibition |
| :---: | :---: | :---: | :---: |
|  |  |  |  |
| $0-4$ | $12.72 \pm 8.58$ | $19.15 \pm 10.77$ | $10.30 \pm 5.23$ |
| $4-8$ | $7.58 \pm 3.48$ | $13.26 \pm 6.89$ | $5.56 \pm 3.26$ |
| $8-10$ | $5.70 \pm 2.89$ | $11.56 \pm 18.00$ | $3.61 \pm 1.63$ |
| $10-24$ | $24.27 \pm 12.78$ | $20.98 \pm 9.74$ | $12.92 \pm 7.65$ |
| $0-24$ | $50.28 \pm 16.90$ | $64.95 \pm 27.93$ | $32.40 \pm 10.97$ |
| $C l_{\text {m } 6(6)}(\mathrm{mL} / \mathrm{min})$ | $0.67 \pm 0.32$ | $0.69 \pm 0.30$ | $0.32 \pm 0.16$ |

[^1]

Figure 3. (A) Correlation between $A U C_{F}$ and CYP3A activity (MR) before inhibition by clarithromycin ( $n=24$ ); (B) Correlation between $A \cup C_{F}$ and CYP3A activity after inhibition by clarithromycin ( $n=24$ ); (C) Correlation between the change in AUC $_{F}$ and CYP3A activity ( $n=24$ ). MR: the plasma concentration ratio of 1-OHMDZ to MDZ at 1 h .

## $6 \beta-0 H F / F$ variations and correlation with CYP3A activity

MDZ administration produced increases in urinary $6 \beta-\mathrm{OHF} / \mathrm{F}$ of $102 \%(P=0.017), 185 \%(P=0.006)$ and $123 \%(P=0.019)$ at $0-4$, $4-8$, and $8-10 \mathrm{~h}$ time-periods, respectively; however, there was no increase in the urinary excretion of cortisol. The values of urinary $6 \beta-\mathrm{OHF} / \mathrm{F}$ at all time-periods decreased significantly when CYP3A was inhibited by clarithromycin, but no changes in the urinary excretion of cortisol were observed (Table 4 and Figure 2). No correlations were found between $6 \beta-\mathrm{OHF} / \mathrm{F}$ and MR, either before or after inhibition $(r=0.063$, $P=0.769 ; r=-0.070, P=0.746$ ) (Figure 5). Furthermore, there was a lack of correlation between the change in $6 \beta-O H F / F$ and MR ( $r=0.001, P=0.995$; Figure 5).

## Effect of MDZ administration on endogenous cortisol levels

No significant changes were found in $\mathrm{AUC}_{\mathrm{F}}$, either before or after MDZ administration ( $P=0.178$; Table 2 and Figure 2). Meanwhile, no significant changes in the amount of urinary $6 \beta$-OHF ( $P=0.169$ ) and F $(P=0.391)$ were measured over the 24 h time-period (Tables 3 and 4).

Table 4. Cortisol urinary excretion and $6 \beta-O H F / F$ variations before and after clarithromycin inhibition. $n=24$. Data are given as mean $\pm$ SD.

| Time period (h) | Cortisol urinary excretion ( $\mu \mathrm{g}$ ) |  |  | 63-OHF/F |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Before inhibition |  | After inhibition | Before inhibition |  | After inhibition |
|  | Non MDZ | With MDZ |  | Non MDZ | With MDZ |  |
| 0-4 | $9.26 \pm 4.37$ | $9.65 \pm 6.71$ | $8.95 \pm 4.58$ | $1.47 \pm 0.82$ | $2.97 \pm 2.46$ | $1.35 \pm 0.79$ |
| 4-8 | $7.91 \pm 4.57$ | $7.69 \pm 5.79$ | $8.02 \pm 6.06$ | $1.20 \pm 0.71$ | $3.44 \pm 3.55$ | $1.43 \pm 2.89$ |
| 8-10 | $4.65 \pm 2.37$ | $4.93 \pm 3.45$ | $4.34 \pm 2.83$ | $1.55 \pm 1.12$ | $3.46 \pm 4.28$ | $1.46 \pm 1.60$ |
| 10-24 | $11.79 \pm 5.05$ | $12.46 \pm 8.72$ | $10.92 \pm 6.87$ | $2.66 \pm 2.36$ | $3.24 \pm 4.14$ | $1.47 \pm 0.87$ |
| 0-24 | $33.61 \pm 12.68$ | $34.72 \pm 15.01$ | $32.24 \pm 13.73$ | $1.71 \pm 0.90$ | $2.27 \pm 1.56$ | $1.17 \pm 0.64$ |

$6 \beta-O H F / F$ : urinary ratio of $6 \beta-O H F / F$.


Figure 4. (A) Correlation between $C I_{m(6 \beta)}$ and CYP3A activity before inhibition by clarithromycin ( $n=24$ ); (B) Correlation between $C I_{m(6 \beta)}$ and CYP3A activity after inhibition by clarithromycin ( $n=24$ ); (C) Correlation between the change in $\mathrm{Cl}_{\mathrm{m}(6 \beta)}$ and CYP3A activity ( $n=24$ ). MR: the plasma concentration ratio of 1-OHMDZ to MDZ at 1 h .

## Discussion

The absence of any correlations between CYP3A activity and $A \cup C_{F}, C I_{m(6 \beta)}$, and $6 \beta-0 H F / F$
The relationships between CYP3A activity and several measures $\left(\mathrm{AUC}_{\mathrm{F}}, C l_{\mathrm{m}(6 \beta)}\right.$, and $\left.6 \beta-\mathrm{OHF} / \mathrm{F}\right)$ both before and after clarithromycin inhibition were examined. However, there were no significant correlations between any of these mea-


Figure 5. (A) Correlation between $6 \beta-O H F / F$ and CYP3A activity before inhibition by clarithromycin ( $n=24$ ); (B) Correlation between 6ß-OHF/F and CYP3A activity after inhibition by clarithromycin ( $n=24$ ); (C) Correlation between the change in $6 \beta-0 H F / F$ and CYP3A activity ( $n=24$ ). MR: the plasma concentration ratio of 1-OHMDZ to MDZ at 1 h .
sures and CYP3A activity (indicated by the secretion and metabolism of cortisol). It is now known that cortisol is produced by the adrenal cortex, and its secretion exhibits a circadian rhythm; the highest levels are secreted between 08:00 and 12:00 and the lowest levels are released at midnight ${ }^{[7]}$. The pathway underlying cortisol metabolism in humans is complex. It has been proposed that the conversions of cortisol
to cortisone and $6 \beta$-hydroxycortisol to $6 \beta$-hydroxycortisone by $11 \beta$-HSD play important roles in cortisol metabolism. The metabolism of cortisone to $6 \beta$-hydroxycortisone is also catalyzed by CYP3A ${ }^{[14]}$. In addition, several other physiological factors might be involved in the disposition of MDZ and F. Firstly, when it is administered orally, MDZ is metabolized not only in the liver but also in the intestine. This affects the accuracy of measurements of $\mathrm{AUC}_{\mathrm{F}}$, as this value mainly reflects hepatic CYP3A activity. Second, according to a previous study, the efflux of P-glycoprotein (P-gp) in vivo probably affects $\mathrm{AUC}_{\mathrm{F}}{ }^{[15]}$. However, as MDZ exhibits characteristics of a class 1 drug in the biopharmaceutical classification system, the disposition of MDZ is influenced to a lesser degree by P-gp ${ }^{[16,17]}$.

Additionally, differences in renal function in different subjects may cause heterogeneity in $6 \beta-\mathrm{OHF}$ excretion that could affect the values of $C l_{m(6 \beta)}$. It is also worth noting that CYP3A consists of at least three isoforms: CYP3A4, CYP3A5, and CYP3A7. Although CYP3A5 may also have played a role here, the relative contribution of each of these isoforms to cortisol metabolism has not been completely elucidated ${ }^{[7]}$.

## $A^{\prime} C_{F}$ is not a suitable index for evaluating variations in CYP3A activity in vivo

Our study reported that there were significant changes in both CYP3A activity (marked by MR) and $A^{\prime} C_{F}$ after clarithromycin treatment in twenty-four healthy subjects. However, in response to a reduction in the values of MR of around $75 \%$, the value of $\mathrm{AUC}_{\mathrm{F}}$ actually increased by $19 \%$; no correlation was found between these measures. This suggested that $A U C_{F}$ was an entirely unsuitable index for evaluating variations in CYP3A activity. As discussed previously, this might be accounted for the regulation of cortisol feedback and physiological factors.

## Inhibitory effect on CYP3A activity in vivo can be partly reflected by $\mathrm{Cl}_{\mathrm{m}(6 \beta)}$ and $6 \beta-0 \mathrm{HF} / \mathrm{F}$

Although no correlations were found between $C l_{\mathrm{m}(6 \beta)}$ and CYP3A activity, or between $6 \beta-\mathrm{OHF} / \mathrm{F}$ and CYP3A activity, significant changes were seen in $C l_{\mathrm{m}(6 \beta)}$ and $6 \beta-\mathrm{OHF} / \mathrm{F}$ with clarithromycin treatment. The reduction in $C l_{\mathrm{m}(6 \beta)}$ of $54.2 \%$ accounted for approximately three-quarters of the decline in MR. Though it did not completely reflect the change in CYP3A activity, it was a more useful marker than $\mathrm{AUC}_{\mathrm{F}}$. This inaccuracy might also be explained by the many factors that influence plasma cortisol levels. Similar to F, $6 \beta$-OHF is also a secretory product of the human adrenal system, and its secretion is thought to be controlled by adrenocorticotrophic hormone ${ }^{[18]}$. Furthermore, individual differences in renal function may affect the degree of $6 \beta$-OHF excretion in different subjects, with a consequent effect on $C l_{\mathrm{m}(6 \beta)}$ values.

A measure of urinary $6 \beta-\mathrm{OHF} / \mathrm{F}$ during the $0-24 \mathrm{~h}$ collection period might also be used to partly evaluate variations in CYP3A activity. After clarithromycin administration, $6 \beta$-OHF/F declined by about $50 \%$. In this study, $6 \beta$-OHF urinary excretion declined by $50 \%$, whereas cortisol excretion did
not vary significantly during the 24 h collection period. This may be due to either decreased cortisol secretion or increased cortisol metabolism, one of which was the augmented conversion of cortisol to cortisone. Thus, measurements of the concentration of cortisone could provide an explanation for why the urinary cortisol level remained stable with clarithromycin pretreatment.

As discussed above, cortisol secretion exhibits a diurnal rhythm. In the present study, the intraindividual variability in measures of $C l_{\mathrm{m}(6 \beta)}$ and $6 \beta$-OHF/F were 7.1- and 11.8 -fold, respectively. This was clearly a major impediment for the establishment of a cortisol-based CYP3A parameter.

## Effect of MDZ administration on endogenous cortisol level

In a previous study ${ }^{[19]}$, the change in plasma cortisol level with MDZ anesthesia was examined in 11 patients that were undergoing abdominal hysterectomy. The plasma cortisol concentration decreased slightly at the induction of anesthesia and at the beginning of surgery. By the end of the surgery, the cortisol levels had increased significantly (the highest value was attained a few hours after the operation), although levels of this hormone approached the normal range on the morning following the operation. Misiolek et al ${ }^{[20]}$ observed the suppression of cortisol release when propofol and MDZ were used for anesthesia induction for non-toxic struma surgery in contrast to thiopentone administration. The results presented here showed the effect of MDZ administration on cortisol levels. With the exception of the plasma cortisol concentration at $1 \mathrm{~h}(25 \%, P=0.033)$, no significant changes were observed at other time points or in the measures of $\mathrm{AUC}_{\mathrm{F}}$ (Table 2). Meanwhile, variations in the amount of urinary $6 \beta$-OHF were detected during the collection periods of $0-4 \mathrm{~h}(51 \%, P=0.022)$, $4-8 \mathrm{~h}(75 \%, P=0.002)$ and $8-10 \mathrm{~h}(103 \%, P=0.019)$, but not in the $10-24 \mathrm{~h}(P=0.160)$, and $0-24 \mathrm{~h}(P=0.169)$ periods. There were also no differences in the $C l_{\mathrm{m}(6 \beta)}$ values during these periods ( $P=0.388$; Table 3). There were no significant changes in the amount of urinary F at any time-period (Table 4). This might be explained by the existence of different mechanisms; it is possible that the reduced plasma cortisol levels at 1 h were mainly due to the enhanced conversion of cortisol to urinary $6 \beta$-OHF or to the substantially elevated F to E conversion which was followed by the transformation of E to $6 \beta$-OHE by CYP3A. This may have resulted in the enhanced conversion of $6 \beta$-OHE to $6 \beta-\mathrm{OHF}$ by 11 -HSD. Because of this complex and slow procedure, the urinary excretion of $6 \beta-\mathrm{OHF}$ during the collection periods of $4-8 \mathrm{~h}$ and $8-10 \mathrm{~h}$ still remained at a high level. Further studies are required to either confirm or reject these hypotheses.

## Conclusion

Although the inhibition of CYP3A activity can be detected by $C l_{\mathrm{m}(6 \beta)}$ and $6 \beta-\mathrm{OHF} / \mathrm{F}$ values, these measures are lacking in accuracy. On the contrary, we have established that $A U C_{F}$ is an entirely inappropriate indicator of changes in CYP3A activity. We have also demonstrated that MDZ administration is without effect on either cortisol metabolism or excretion over
a 24 h time period.

## Acknowledgements

The work was supported by the Health Department of Hu-nan Province.

## Author contribution

Ze-neng CHENG designed research; Xiao-min LI and Zhe-yi HU performed research; Xi LUO wrote the paper.

## References

1 Williams JA, Ring BJ, Cantrell VE, Jones DR, Eckstein J, Ruterbories K, et al. Comparative metabolic capabilities of CYP3A4, CYP3A5, and CYP3A7. Drug Metab Dispos 2002; 30: 883-91.
2 Krishna DR, Shekar MS. Cytochrome P450 3A: genetic polymorphisms and inter-ethnic differences. Methods Find Exp Clin Pharmacol 2005; 27: 559-67.
3 Dahl ML. Cytochrome P450 phenotyping/genotyping in patients receiving antipsychotics: useful aid to prescribing? Clin Pharmacokinet 2002; 41: 453-70.
4 Zaigler M, Tantcheva-Poor I, Fuhr U. Problems and perspectives of phenotyping for drug-metabolizing enzymes in man. Int J Clin Pharmacol Ther 2000; 38: 1-9.
5 Streetman DS, Bertino JS, Nafziger AN. Phenotyping of drugmetabolizing enzymes in adults: a review of in vivo cytochrome P450 phenotyping probes. Pharmacogenetics 2000; 10: 187-216.
6 Zhu B, Ou-Yang DS, Cheng ZN, Huang SL, Zhou HH. Sing plasma sampling to predict oral clearance of CYP3A probe midazolam. Acta Pharmacol Sin 2001; 22: 634-38.
7 Galteau MM, Shamsa F. Urinary 6ß-hydroxycortisol: a validated test for evaluating drug induction or drug inhibition mediated through CYP3A in humans and in animals. Eur J Clin Pharmacol 2003; 59: 713-33.
8 Chen YC, Gotzkowsky SK, Nafziger AN, Kulawy RW, Rocci ML, Bertino JS, et al. Poor correlation between 6-hydroxycortisol: cortisol molar ratios and midazolam clearance as measure of hepatic CYP3A activity. Br J Clin Pharmacol 2006; 62: 187-95.
9 Kinirons MT, O Shea D, Downing TE, Fitzwilliam AT, Joellenbeck L, Groopman JD, et al. Absence of correlations among three putative in vivo probes of human cytochrome P4503A activity in young healthy
men. Clin Pharmacol Ther 1993; 54: 621-9.
10 Hunt CM, Watkins PB, Saenger P, Stave GM, Barlascini N, Watlington CO, et al. Heterogeneity of CPY3A isoforms metabolizing erythromycin and cortisol. Clin Pharmacol Ther 1992; 51: 18-23.
11 Furuta T, Suzuki A, Mori C, Shibasaki H, Yokokawa A, Kasuya Y. Evidence for the validity of cortisol $6 \beta$-hydroxylation clearance as a new index for in vivo cytochrome P450 3A phenotyping in humans. Drug Metab Dispos 2003; 31: 1283-7.
12 Hu Z, Gong Q, Hu X, Wang L, Cao Y, Cao W, et al. Simultaneous determination of $6 \beta$-hydroxycortisol and cortisol in human urine and plasma by liquid chromatography with ultraviolet absorbance detection for phenotyping the CYP3A activity. J Chromatogr B 2005; 826: 238-43.
13 Eeckhoudt SL, Desager JP, Horsmans Y, De Winne AJ, Verbeeck RK. Sensitive assay for midazolam and its metabolite 1'-hydroxymidazolam in human plasma by capillary high-performance liquid chromatography. J Chromatogr B 1998; 710: 165-71.
14 Abel SM, Maggs JL, Back DJ, Park BK. Cortisol metabolism by human liver in vitro-l. Metabolite identification and inter-individual variability. J Steroid Biochem Mol Biol 1992; 43: 713-9.
15 Ueda K, Okamura N, Hirai M, Tanigawara Y, Saeki T, Kioka N, et al. Human P-glycoprotein transports cortisol, aldosterone, and dexamethazone, but not progesterone. J Biol Chem 1992; 267: 24248-52.
16 Tolle-Sander S, Rautio J, Wring S, Polli JW, Polli JE. Midazolam exhibits characteristics of a highly permeable P-glycoprotein substrate. Pharm Res 2003; 20: 757-64.
17 Custodio JM, Wu CY, Benet LZ. Predicting drug disposition, absorption/elimination/transporter interplay and the role of food on drug absorption. Advanced Drug Delivery Rev 2008; 60: 717-33.
18 Szucs N, Varga I, Patocs A, Toth M, Glaz E, Racz K. Secretion of $6 \beta$-hydroxycortisol by normal human adrenals and adrenocortical adenomas. Steroids 2003; 68: 477-82.
19 Kertesz A, Godo G, Falkay G, Boros M. Plasma cortisol, prolactin and thyroxine levels related to midazolam anaesthesia. Acta Med Hung 1986; 43: 283-9.
20 Misiolek H, Wojcieszek E, Dyaczynska-Herman A. Comparison of influence of thiopentone, propofol and midazolam on blood serum concentration of noradrenaline and cortisol in patients undergoing non-toxic struma operation. Med Sci Monit 2000; 6: 319-24.


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    Received 2009-04-23 Accepted 2009-06-24

[^1]:    $\mathrm{Cl}_{\mathrm{m}(6 \beta)}$ : cortisol $6 \beta$-hydroxylation clearance.

